Yeast Biocapsules: More than just a carrier for food flavours

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INTRODUCTION



- What has yeast ever done for us?
- Properties of yeast
- Target molecules process improvements
- Future prospects

What Has Yeast Ever Done For Us?

 Over 5000 years ago yeast was used in fermentation for wine and bread-making





Pure yeast used in brewing
 from 1883 by Carlsberg's
 Emil Hansen

• Yeast cell factories



Good Yeast vs Bad Yeast

Baker's and Brewer's yeast
 Saccharomyces cerevisiae or
 S. pastorianus (aka S. carlsbergensis)

- Food and drink spoilage caused by "wild yeast" and moulds.
- Yeast Infections (Candidiasis)



Biocapsules



• Yeast cells as natural capsules for functional ingredients



Why Use Yeast As An Inert Carrier?

- Pre-formed microcapsules
- Produces readily dispersible dry powders and granules
 - Spray drying or spray agglomeration
- Contents are protected by a robust cell wall
 - Products are amenable to blending, extrusion and high heat processing
- Ideal for small fat-soluble molecules (hydrophobes)
- Yeast is a readily available natural raw material, a commodity product with a consistent supply









Target Ingredients

• Flavours and tastes

- Flavour and aroma
- Cooling agents
- Health and wellness ingredients

Agriculture

- Fungicides
- Insecticides and herbicides
- Semiochemicals/attractants

Healthcare

- Pharmaceutical APIs & Paracellular drug delivery
- Repellents
- Wound care, sanitizers, cleaning products

Same Old Challenges



- Convert liquids to solids Improve handling
- Targeted delivery Improve impact or bioavailability
- Controlled & delayed release Process stability
- Masking taste and odour bitter plant extracts and volatiles
- Protection for sensitive ingredients against
 - VU 👯

🍟 Heat





- Isolation of reactive components
- Stabilisation of volatile ingredients to improve shelf life
 - Flavours and fragrances

Production - Spray Drying





- Emulsifiable liquids such as essential oils and liquid flavours
- Typically water or oil soluble principal components are typically mixed with a matrix material e.g. maltodextrin and gums or <u>yeast</u>
- A two-fluid nozzle disperses the liquid into fine droplets, water evaporates in the drying chamber forming dry particles

Production - Spray Drying





Uptake Profile Of Tea Tree Oil



When yeast was used to encapsulate antimicrobial Tea Tree oil the initial rate of uptake was rapid over the first 30 minutes and approached a plateau after 5 hours

Duckham et al. Proc. of 14th International Symposium on Microencapsulation, September 4-6 2003.

Localisation Within Cells



Fluorescence microscopy



Freeze fracture scanning electron microscopy



Confocal-fluorescence and scanning electron microscopy used to visualise encapsulation Nile red (in green) to indicate lipid droplets and DAPI stain to show the nucleus (in blue)

Mechanism Of Encapsulation



After Dardelle et al. (2007). Food Hydrocolloids. 21, 953-960

Unique Barrier Protection Of Cell Contents



The structure of the yeast protective envelope comprises the cell wall and the lipid membrane, the key selective barriers in the uptake of fat soluble flavours



- Nile Red is non-ionic and emission does not depend on local pH, or on the presence of specific chemical compounds
- It has poor water solubility and exhibits bright red-yellow fluorescence in a hydrophobic environment
- Exhibits solvatochromic behaviour



NR ideal for targeting intracellular lipidic droplets

Staining With Nile Red



NR exhibits negligible fluorescence in water and fluoresces according to the medium it is in. Due to these useful spectral properties it has been extensively used for microscopic imaging purposes; in particular for intracellular lipidic droplets



after Ciamponi, et al. (2012) Applied Microbiol Biotechnol. 95, (6) pp 1445-1456

Visualisation Of Absorbed Fat Soluble Flavour



Appl Microbiol Biotechnol DOI 10.1007/s00253-012-4127-8

BIOTECHNOLOGICAL PRODUCTS AND PROCESS ENGINEERING

Yeast cells as microcapsules. Analytical tools and process variables in the encapsulation of hydrophobes in *S. cerevisiae*

Federica Ciamponi · Craig Duckham · Nicola Tirelli

Received: 24 January 2012 / Revised: 20 April 2012 / Accepted: 20 April 2012 © Springer-Verlag 2012

Abstract Yeast cells can be used as biocompatible and biodegradable containers for the microencapsulation of a variety of actives. Despite the wide application of this process, e.g. in the food industry, mechanism and controlling factors are yet poorly known. In this study we have studied kinetics and mechanistic aspects of the spontaneous internalization of terpenes (as model hydrophobic compounds) in *Saccharomyces cerevisiae*, quantifying their encapsulation through HPLC analysis and

Electronic supplementary material The online version of this article (doi:10.1007/s00253-012-4127-8) contains supplementary material, which is available to authorized users. fluorescent staining of lipidic bodies with Nile Red, while in parallel monitoring cell viability. Our results showed that this encapsulation process is essentially a phenomenon of passive diffusion with negligible relevance of active transport. Further, our evidence shows that the major determinant of the encapsulation kinetics is the solubility of the hydrophobe in the cell wall, which is inversely related to partition coefficient (log *P*).

Keywords Encapsulation · Yeast · Cell wall · Flavours · Diffusion

Hydrophobic Probes



• Early studies showed that components of garlic oil were encapsulated with high efficiency

 Polymer probes were created to investigate compartmentalisation and test the molecular weight cut off characteristics of yeast biocapsules

Fluorescent Probes Synthesised For The Study



The dansyl group was chosen for its high stability both to chemical agents and to photo-bleaching (λ_{exc} = 343 nm; λ_{em} = 494 nm)

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Kilcher et al. (2008) Faraday Discuss. 139, 199-212

Cell Membrane Removal

S. cerevisiae yeast cells initially in PBS, are transferred into DMSO for 2hr and then spun down and returned to PBS

A 20 nm thick gap appears where the lipid bilayer resided between the cytoplasm and cell wall, initially clearly visible, seems to disappear, as do the membranes of internal cell organelles. These membranes are not restored when cells are transferred back to water milieu after exposure to DMSO.



Improvements In The Uptake Process

Conventional Approach + 0.5 mM probe

Conventional Approach following DMSO pre-treatment

New Solvent Based Method after Kilcher et al 2008



This shows how droplets of fluorescent polysulfides above 600 daltons could not pass across the cell wall using conventional aqueous mixing. Selected solvents in a water free system facilitated the process for very hydrophobic molecules to almost 4000 daltons.

Kilcher et.al. (2008) Faraday Discuss. 139, 199-212

Testing Yeast Viability During Encapsulation

Appl Microbiol Biotechnol DOI 10.1007/s00253-012-4127-8

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Images of the localization and fluorescence of FUN-1 Cells were also stained 25 μ M Calcofluor white to highlight the cell wall.

Ciamponi, et al. (2012) Applied Microbiol Biotechnol. 95, (6) pp 1445-1456

Raw Materials - Dead Yeast



Overview

- Trigger:
 - Moisture
 - pH
 - Temperature
 - Pressure
 - Shear

Release:

- Sustained Diffusion
- Burst
 Shear rupture

Target:

- Crop protection Fungicide/Herbicide/Insecticide
- APIs
 - Oral/Topical/Paracellular DD
- Flavours





BENEFITS: Improved Delivery



- Effective delivery
 - Modified flavour profiles
 - Improved bioavailability
 - A sustained release platform delivery system

Protection from

- Evaporation
- High temperatures
- High shear processes
- Structurally from pH changes and hydrolysis
- Prospects for new product concepts
- Stable aqueous dispersions for fat soluble ingredients
- Easy-to-handle: fine powder and liquid spray coatings
- Natural retain ingredient labelling advantages

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